



Diversity of fungal endophytes in leaves of *Mentha piperita* and *Mentha canadensis*

Leonardo Wedderhoff Herrmann¹, Carolina Gracia Poitevin², Luzianna Celeste Schuindt³,
Angela Bozza De Almeida⁴, Ida Chapaval Pimentel⁵

¹⁻⁵ Basic Pathology Department, Federal University of Paraná, Curitiba, Paraná, Brazil

Abstract

Endophytic fungi are microorganisms that colonize internal plant tissues without causing immediate negative effects, being present in virtually the entire plant kingdom. Despite this, much of its biotechnological potential is unknown, and its identification is still recent. Peppermint, popular name of the genus *Mentha*, is a perennial plant native of Europe with various applications in cosmetics, pharmaceuticals, food and hygiene products. The objective of this work was the isolation and identification of endophytic fungi from leaves of two peppermint species, *Mentha piperita* and *Mentha canadensis*. Identification was performed by macro and micromorphology and by the sequencing of the ITS1-5.8S-ITS2 region of ribosomal DNA and partial β -tubulin gene. Thirty-five endophytes were identified in total, belonging to the genus *Alternaria*, *Aspergillus*, *Curvularia*, *Epicoccum*, *Fusarium*, *Penicillium*, *Periconia* and *Phoma* and the order *Xylariales*. This is the first report of endophytic fungi diversity in *M. canadensis*, and provides knowledge on their future use on biotechnological processes.

Keywords: peppermint, isolation, endophytic

1. Introduction

Endophytic microorganisms are organisms that colonize the interior of plants without causing any visible negative effects at first, being present in practically all plant species [1]. Although it is a propagated concept, there are still doubts about the definition of endophytic, wherein some of which may be latent pathogens or cause damage to the plant [2,3]. However, the great majority presents themselves as microorganisms that develop in the apoplasmic fluid of the plants in an asymptomatic way, acting commonly in a mutualistic way and often increasing the resistance of the plant against drought and diseases, facilitating its growth. Thus, the definition of endophyte has been extended to include all microorganisms that colonize internal tissues of plants, either for benefit, detriment or in a neutral way [2,4,5]. Potentially promising, endophytic microorganisms can be used as sources of primary metabolites [6] and secondary metabolites. Some advantages are attributed to the presence of endophytes, such as the ability to produce antibiotics [7] and other secondary metabolites of pharmacological interest; employment as agents of biological control of diseases and pests (bio-pesticides) [8]; bioremediation of soil contaminated with pollutants; and use as vectors for introducing genes into host plants [3,5,9].

The peppermint, as it is popularly known the genus *Mentha*, is an eudicotyledone plant which belongs to the *Lamiaceae* family, cultivated by the ancient Egyptians and documented since before the thirteenth century [10]. It is a perennial medicinal plant, native of Europe and naturalized in several parts of the world, and it is often used on a large scale as a component of products intended for cosmetic and pharmaceutical use [11]. In addition to its high morphological variability, most peppermint species are characterized by their high chemical diversity in terms of the constituents present in their leaves [12]. The peppermint essential oil is composed mainly of mentone, menthol, eucalyptol, linalool

and alpha-terpinene, and it has antibacterial activity against *Candida albicans*, *Escherichia coli*, *Salmonella choleraesuis* and *Staphylococcus aureus* [13].

Its properties are well known in traditional medicine, and its antimicrobial, anti-viral, antifungal, anti-oxidant and anti-allergic characteristics have been tested and proven, either as essential oil [10,14], aqueous extract [12,15] or tea [11]. There are reports in the literature about tests performed with essential oil of peppermint to verify its effectiveness against the growth of *Aspergillus niger* and *A. carbonarius*, ochratoxin A producers [16], and *A. flavus*, aflatoxin producer [17]. All these properties may associate peppermint to a possible source of endophytic fungi with biotechnology capability. Therefore, the objective of this work was to isolate endophytic fungi from leaves of two peppermint species, *Mentha piperita* and *Mentha canadensis*, and to identify them by molecular techniques.

2. Materials and methods

2.1 Isolation of endophytic fungi

The endophytic fungi were isolated from healthy peppermint leaves (*M. piperita* and *M. canadensis*) collected at the Canguiri Experimental Station Center of the Federal University of Paraná (UFPR) (25°23'30"S and 49°07'30" W), located in the municipality of Pinhais, PR, Brazil. The leaves were initially washed with water to remove soil and dust and then dipped in 70% alcohol for 30 seconds and 1% sodium hypochlorite for 3 minutes [18]. Subsequently, the material was washed twice in sterile distilled water and cut into 4 mm² fragments, and 5 fragments were placed in each Petri dish (90 x 15 mm) containing potato dextrose agar (PDA) with tetracycline (100 μ L mL⁻¹). The abaxial part of the leaves was in contact with the medium. The plates were placed in a BOD incubator at 28 \pm 1 °C for 7 days. The grown fungi were isolated in Petri dishes containing PDA medium and the isolation frequency was calculated [19].

2.2 Morphological characterization

Single-spore colonies were cultivated in PDA and incubated at 28 ± 1 °C for 14 days. The isolates were separated into morphogroups according to their macro and micromorphological characteristics. The isolates are deposited in the Microbiological Collection of Paraná Network - Taxonline (CMRP), Basic Pathology Department, Federal University of Paraná, Curitiba, Brazil.

2.3 Molecular identification of endophytic fungi

For molecular identification, the ITS1-5,8S-ITS2 region of ribosomal DNA and partial β -tubulin gene were sequenced. The DNA was extracted according to Badali [20], and PCR was made using the pairs of primers ITS1 and ITS4 [21] and Bt2a and Bt2b [22]. Sequencing was performed by the chain

termination method [23], in ABI 3500 automatic DNA sequencer (Applied Biosystems). The sequences were aligned and edited with the Staden program version 1.6 [24], aligned with the MEGA 7 program [25] and compared with other sequences in the NCBI database by the BLAST program [26].

3. Results

From *M. piperita*, 220 fungi belonging to 50 different morphogroups were isolated, and the isolation frequency was 40.8%. For *M. canadensis*, 73 fungi belonging to 19 morphogroups were isolated, with a frequency of 14.6%. In total, 33 fungi were identified from the ITS region and 2 from partial β -tubulin gene sequencing (TABLE 1), being 10 of *M. canadensis* and 25 of *M. piperita*.

Table 1: Endophytic fungi isolated from leaves of *Mentha canadensis* and *Mentha piperita*, identified by the ITS region and partial β -tubulin gene sequencing.

Host	Identification	Frequency (%)	Isolate	Genbank accession number	Homology (%)
<i>Mentha canadensis</i>	<i>Alternaria alternata</i>	67,7	100	MK370625	100
			92-E1	MK370626	100
			83-D1	MK370627	100
			11-B1	MK370628	100
			41-B1	MK370629	99
			28-C1	MK370630	100
	<i>Aspergillus fumigatus</i>	9,7	70-E1	MK370631	100
	<i>Curvularia trifolii</i>	12,9	89-E1	MK370632	100
<i>Mentha piperita</i>	<i>Fusarium equiseti</i>	3,2	74-D1	MK370633 / MK373039	100
	<i>Phoma</i> sp.	6,5	65-E1	MK370634	100
	<i>Alternaria alternata</i>	52,8	75-D1	MK370635	100
			20-A1	MK370636	100
			63-A1	MK370637	100
			86-A1	MK370638	100
			93-D1	MK370639	100
			94-C1	MK370640	100
			96-C1	MK370641	100
			47-A2	MK370642	99
	<i>Alternaria solani</i>	8,7	61-C1	MK370643	100
			52-B1	MK370644	99
			91-B1	MK370645	99
	<i>Alternaria</i> sp.	23,6	36-C1	MK370646	100
			14-B1	MK370647	100
<i>Aspergillus nomius</i>	1,2	57-D1	MK370648	99	
<i>Curvularia trifolii</i>	0,6	89-B1	MK373042	99	
		50-C1	MK373043	99	
		64-C1	MK370649	100	
		09-B1	MK370650	100	
		31-E1	MK370651 / MK373040	100	
		95-A1	MK370652 / MK373041	100	
		86-B1	MK370653	100	
		39-A1	MK370654	99	
		31-C1	MK370655	99	
		94-E1	MK370656	99	
<i>Periconia byssoides</i>	3,1	03-D1	MK370657	98	
		<i>Xylariales</i> sp.	7,5		

4. Discussion

This is the first study of endophytic fungi isolated from leaves of *M. canadensis* and the first record of the genus *Curvularia* and *Periconia* and the order *Xylariales* in *M. piperita*. The identified endophytic fungi were *Alternaria*, *Aspergillus*, *Curvularia*, *Epicoccum*, *Fusarium*, *Penicillium*, *Periconia*, *Phoma* and *Xylariales*.

The genus *Colletotrichum*, *Drechslera*, *Fusarium*, *Guignardia*, *Nigrospora* and *Xylaria* were isolated from *Mentha spicata* [27], among them only the genus *Fusarium* and the order *Xylariales* were identified in this paper in the

leaves of *M. piperita* and *M. canadensis*. Another study reported the presence of an endophytic ascomycete isolate in stems and leaves of *M. piperita* without causing damage to the host plant [28], which may indicate interactions similar to those of the fungi identified in this study.

Zimowska [29] isolated endophytes from several parts of the *M. piperita* plant, and identified the genus *Alternaria*, *Epicoccum*, *Fusarium*, *Penicillium* and *Phoma* in common with the present paper, as well as other genus such as *Cladosporium* and *Trichoderma*. Also in this study, the genus *Phoma* was identified as a possible cause of necrosis. In

previous studies, Zimowska [30] reports that *Phoma* and *Fusarium* species were less frequent in peppermint leaves, wherein the *Alternaria alternata* species was predominant, which is in agreement with our results.

In a study with *M. piperita* samples from several regions of India, it was identified 63 endophytes. The genus in common with the present study are *Alternaria*, *Aspergillus*, *Fusarium* and *Penicillium*, emphasizing again the species *Alternaria alternata* [31].

The genus of fungi presented in this paper have been isolated from several species of plants, however studies with peppermint are still scarce. The species *A. alternata* was isolated from coffee and presented antimicrobial and antioxidant activity [32]. Lin [33] isolated 174 endophytic fungi from *Camptotheca acuminata*, a medicinal plant, with 12.6% of *Alternaria* sp. This genus was also isolated from tangerine, lemon [34], soybean [35], eucalyptus [36], *Crambe* sp. [37], peach palm [38], tomato, arugula, cauliflower, wheat, grape, apple, among others [39]. It is important to emphasize that the presence of *Alternaria* spp. in peppermint leaves may be an indicator of the presence of mycotoxins such as alternariol, alternariol monomethyl ether, tenuazonic acid and altertoxins [40].

In addition to *Alternaria*, other endophytic fungi from peppermint were the genus *Epicoccum*, *Curvularia* and *Periconia*, as well as the order *Xylariales*. The fungus *Epicoccum nigrum* has been isolated from peach palm (*Bactris gasipaes*) [38], rubber tree [41], maize [42], cabbage [43] and olive [44]. Madrigal and Melgarejo [45] showed that this species has antifungal activity, producing compounds such as flavipin.

The genus *Curvularia* is associated with plant diseases, such as leaf spot [46], and humans diseases [47]. The same genus has already been isolated from dwarf papyrus sedge, *Cenchrus ciliaris* [48], ginger [49], cacao [50], carrot [51], *Lippia sidoides* [52] and rice [53]. *Hypoxylon*, one of the genus that belong to *Xylariales*, can be found as an endophytic of winter cherry [54] and *Persea indica*, producing volatile compounds with potential as biofuel [55,56].

Two antitumor substances (pericosines A and B), effective against leukemia cells, have been isolated from *Periconia byssoides* extracts [57]. The endophytic genus was also found in medicinal plants such as *Lippia sidoides* [52], *Vitex negundo* [58], bear's breeches (*Heracleum sosnowskyi*) [59], ginger [49], *Ougeinia ojeinensis* [60] and grasses [61].

The genus *Fusarium*, found in this study, is also considered a phytopathogen and producer of secondary metabolites toxic for humans and animals [62, 63]. Species of this genus were isolated from barley, wheat, onion, beet, straw [62], plane trees, *Artemisia annua*, winter cherry [54], rubber tree [41], maize [64], *Apodytes dimidiata* [65] and peach palm [38].

Another producer of mycotoxins is the genus *Phoma*, associated with the leaf spot of *Phaeosphaeria*, a disease that attacks tropical and subtropical maize producing areas [66]. Some species have already been isolated from mangrove trees [67], *Cyperus laevigatus*, *Chloris inflata* [48], *Fallopia japonica* [68], maize [42], carrot [51], rubber tree [41], and baobab [69].

It is important to emphasize that the species *Penicillium crustosum*, producer of penitrem toxin A [70], was first identified in peppermint. Different *Penicillium* species have already been found in mangrove trees [67, 71], rubber tree [41], *Pappophorum krapovickasii* [48], African oil palm [72], *Vitex negundo* [58] and cabbage [43], evidencing the diversity of the

genus in plants.

There are no records in the literature of the presence of *Aspergillus fumigatus* in peppermint plants. This genus, isolated from mangrove, has been used in the production of drugs, such as lovastatin [73] and as phosphate solubilizers [71]. Besides, it has been isolated from coffee beans [74, 75], dwarf papyrus sedge [48], African oil palm [72], *Vitex negundo* [58] and *Anemopsis californica* [76].

5. Conclusions

This is the first report of endophytic fungi diversity in *M. canadensis*, and the presence of the genus *Curvularia* and *Periconia* and the order *Xylariales* in *M. piperita*, which provides knowledge on their future use on biotechnological processes.

6. Acknowledgment

The author thanks the National Council for Scientific and Technological Development (CNPq), in the modality of Institutional Program for Scientific Initiation Scholarships (PIBIC), for the financial support.

7. References

- Schulz B, Boyle C. The endophytic continuum. *Mycological Research*. 2005; 661-686.
- Sikora RA, Schäfer K, Dababat AA. Modes of action associated with microbially induced *in planta* suppression of plant-parasitic nematodes. *Australasian Plant Pathology*. 2007; 36:124-134.
- Hyde KD, Soyong K. The fungal endophyte dilemma. *Fungal diversity*. 2008; 33:163-173.
- Sieber TN. Endophytic fungi in forest trees: are they mutualists? *Fungal biology reviews*. 2007; 21:75-89.
- Dutta D, Puzari KC, Gogoi R, Dutta P. Endophytes: exploitation as a tool in plant protection. *Agriculture, Agribusiness and Biotechnology*. Braz. arch. biol. technol. 2014; 57:5.
- Stamford TL, Araújo JM, Stamford NP. Atividade enzimática de microrganismos isolados de jacatupé (*Pachyrhizus erosus* L. Urban). *Ciência e Tecnologia dos Alimentos*. 1998; 18:382-385.
- El-Shatoury S, Abdulla H, El-Karaaly O, El-Kazzaz W, Dewedar A. Bioactivities of Endophytic Actinomycetes from Selected Medicinal Plants in the World Heritage Site of Saint Katherine, Egypt. *Internacional Journal of Botany*. 2006; 2(3):307-312.
- Santos LS, Oliveira MN, Guilhon GMSP, Santos AS, Ferreira ICS, Lopes-Júnior ML, *et al.* Potencial Herbicida da Biomassa e de Substâncias Químicas Produzidas pelo Fungo Endofítico *Pestalotiopsis guepinii*. *Planta Daninha*. 2008; 26(3):539-548.
- Backman PA, Sikora RA. Endophytes: an emerging tool for biological control. *Biological Control*. 2008; 46:1-3.
- Singh R, Shushni MAM, Belkheir A. Antibacterial and antioxidant activities of *Mentha piperita* L. *Arabian Journal of Chemistry*. 2015; 8(3):322-328.
- Mckay DL, Blumberg JBA. Review of the bioactivity and potential health benefits of peppermint tea (*Mentha piperita* L.). *Phytotherapy Research*. 2006; 20:619-633.
- Rosal LF, Leite CD, Maia AJ, Faria CMDR, Baldin I, Marcondes MM, *et al.* Avaliação *In Vitro* do uso do extrato aquoso de hortelã em diferentes concentrações sobre o crescimento micelial do *Penicillium* sp. *Revista Brasileira de Agroecologia*. 2009; 4:2.

13. Silva LF, Cardoso MG, Batista LR., Gomes MS, Rodrigues LMA, Rezende DACS, *et al.* Chemical characterization, antibacterial and antioxidant activities of essential oils of *Mentha viridis* L. and *Mentha pulegium* L. American Journal of Plant Sciences. 2015; 6(5):666-675.
14. Arruda TA, Antunes RMP, Catão RMR, Lima EO, Sousa DP, Nunes XP, *et al.* Preliminary study of the antimicrobial activity of *Mentha x villosa* Hudson essential oil, rontundifolone and its analogues. Revista Brasileira de Farmacognosia. 2006; 16(3):307-311.
15. Venturoso LR, Bacchi LMA, Gavassoni WL. Atividade antifúngica de extratos vegetais sobre o desenvolvimento de fitopatógenos. Summa Phytopathologica. 2011; 37(1):18-23.
16. Passone MA, Girardi NS, Etcheverry M. Evaluation of the control ability of five essential oils against *Aspergillus* section Nigri growth and ochratoxin A accumulation in peanut meal extract agar conditioned at different water activities levels. International Journal of Food Microbiology. 2012; 159:198-206.
17. Dwivedy AK, Prakash B, Chanotiya CS, Bisht D, Dubey NK. Chemically characterized *Mentha cardiaca* L. essential oil as plant based preservative in view of efficacy against biodeteriorating fungi of dry fruits, aflatoxin secretion, lipid peroxidation and safety profile assessment. Food and Chemical Toxicology. 2017; 106(A):175-184.
18. Araújo WL, Lima AOS, Azevedo JL, Marcon J, Kuklinsky-Sobral J, Lacava PT. Manual: Isolamento de microorganismos endofíticos Calq, Piracicaba. 2002.
19. Azevedo JL. Microorganismos endofíticos. In: I.S. MELOJ and L. AZEVEDO, eds. Ecologia Microbiana. Jaguariúna: Editora Embrapa. 1998; 1:117-137.
20. Badali H, Carvalho VO, Vicente V, Attili-Angelis D, Wiatkowski IB, Ende AHGGVD, *et al.* *Cladophialophora saturnica* sp. nov., a new opportunistic species of *Chaetothyriales* revealed using molecular data. Medical Mycology. 2009; 47:55-66.
21. White TJ, Bruns T, Lee S, Taylor J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. Apud Innis MA, Gelfand DH, Shinsky JJ, White TJ, editors. San Diego: PCR Protocols: A Guide to Methods and Applications, Academic Press. 1990; 315-322.
22. Glass NL, Donaldson GC. Development of primer sets designed for use with the PCR to amplify conserved genes from filamentous ascomycetes. Applied and Environmental Microbiology. 1995; 61(4):1323-1330.
23. Sanger F, Nicklen S, Coulson AR. DNA sequencing with chain-terminating inhibitors. Washington: Proceedings of the National Academy of Sciences of the United States of America. 1977; 74:5463-5467.
24. Bonfield J, Beal K, Jordan M, Chen Y, Staden R. The Staden Package Manual. Cambridge: Medical Research Council. 2002.
25. Kumar S, Stecher G, Tamura K. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. – Molecular Biology Evolution. 2016; 33:1870-1874.
26. Altschul SF, Madden TL, Schaffer AA, Zhahg J, Zhang Z, Miller W, *et al.* Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Oxford: Nucleic Acids Research. 1997; 25:3389-3402.
27. Correia JMM, Neves RO, Fraccannabia NR, Ribeiro IATA, Santos IP, Cavalcanti MS. Atividade amilolítica de fungos endofíticos isolados de *Mentha spicata* L. Relatório de reunião, 65ª reunião anual da SBPC, 2014.
28. Mucciarelli M, Scannerini S, Berteza C M, Maffei M. An ascomycetous endophyte isolated from *Mentha piperita* L.: biological features and molecular studies. Mycologia. 2002; 94(1):28-39.
29. Zimowska, B. Diversity of fungi occurring on herbs from *Lamiaceae* family. Phytopathologia. 2010; 56:5-15.
30. Zimowska B. Fungi colonizing and damaging different parts of peppermint (*Mentha piperita* L.) cultivated in South-Eastern Poland. Herba Polonica. 2007; 53(4).
31. Chowdhary K, Kaushik N. Biodiversity study and potential of fungal endophytes of peppermint and effect of their extract on chickpea rot pathogens. Archives of Phytopathology and Plant Protection. 2018; 51(3-4):139-155.
32. Fernandes MRV, Silva TAC, Pfenning LH, Costa-Neto CMda, Heinrich TA, Alencar SMde, *et al.* Biological activities of the fermentation extract of the endophytic fungus *Alternaria alternata* isolated from *Coffea arabica* L. Braz. J. Pharm. Sci. 2009; 45:4.
33. Lin X, Lu C, Huang Y, Zheng Z, Su W, Shen Y. Endophytic fungi from a pharmaceutical plant, *Camptotheca acuminata*: isolation, identification and bioactivity. World J. Microbiol. Biotechnol. 2007; 23:1037-1040.
34. Dini-Andreote F, Pietrobon VC, Dini-Andreote F, Romão AS, Spósito MB, Araújo WL. Variabilidade genética de isolados brasileiros de *Alternaria alternata* por meio de marcadores moleculares de AFLP e RAPD. Brazilian Journal of Microbiology. 2009; 40(3):670-677.
35. Oviedo MS, Ramirez ML, Barros GG, Chulze SN. Impact of water activity and temperature on growth and alternariol and alternariol monomethyl ether production of *Alternaria alternata* isolated from soybean. Journal of Food Protection. 2010; 2:212-404.
36. Javaid A, Samad S. Screening of allelopathic trees for their antifungal potential against *Alternaria alternata* strains isolated from dying-back *Eucalyptus* spp. Natural Product Research: Formerly Natural Product Letters. 2012; 18:26.
37. Carneiro SMdeTPG, Romano E, Marianowski T, Oliveira JP, Garbim THS, Araújo PM. Ocorrência de *Alternaria brassicicola* em crame (*Crambe abyssinica*) no estado do Paraná. Botucatu: Summa Phytopathologica. 2009; 35:2.
38. Almeida CV de, Yara R, Almeida M de. Fungos endofíticos isolados de ápices caulinares da pupunheira cultivada *in vivo* e *in vitro*. Pesquisa agropecuária brasileira. 2005; 40:5.
39. Patriarca A. *Alternaria* in food products. Current Opinion in Food Science. 2016; 11:1-9.
40. Tralamazza SM, Piacentini KC, Iwase CHT, Rocha L. O. Toxicogenic *Alternaria* species: impact in cereals worldwide. Current Opinion in Food Science. 2018; 23:57-63.
41. Gazis R, Chaverri P. Diversity of fungal endophytes in leaves and stems of wild rubber trees (*Hevea brasiliensis*) in Peru. Fungal Ecology. 2010; 3(3):240-254.
42. Sørensen JL, Aveskamp MM, Thrane U, Andersen B. Chemical characterization of *Phoma pomorum* isolated

- from Danish maize. *International Journal of Food Microbiology*. 2010; 136(3):310-317.
43. Park JY, Oyaizu H, Okada G, Takahashi M. Screening of fungal antagonists against yellows of cabbage caused by *Fusarium oxysporum* f. sp. *conglutinans*. *Mycoscience*. 2002; 43(6):447-451.
 44. Preto G, Martins F, Pereira JA, Baptista P. Fungal community in olive fruits of cultivars with different susceptibilities to anthracnose and selection of isolates to be used as biocontrol agents. *Biological Control*. 2017; 110:1-9.
 45. Madrigal C, Melgarejo P. Morphological effects of *Epicoccum nigrum* and its antibiotic flavipin on *Monilia laxa*. *Can. J. Bot.* 1994; 73:425-431.
 46. Aguas Y, Hincapie M, Fernández-Ibáñez P, Polo-López MI. Solar photocatalytic disinfection of agricultural pathogenic fungi (*Curvularia* sp.) in real urban wastewater. *Science of The Total Environment*. 2017; 607-608: 1213-1224.
 47. Gao JX, Chen J. Involvement of a Polyketide Synthetase CIPKS18 in the Regulation of Vegetative Growth, Melanin and Toxin Synthesis, and Virulence in *Curvularia lunata*. *Plant Pathol J*. 2017; 33(6): 597-601.
 48. Loro M, Valero-Jiménez CA, Nozawa S, Márquez LM. Diversity and composition of fungal endophytes in semiarid Northwest Venezuela. *Journal of Arid Environments*. 2012; 85:46-55.
 49. Ginting RCB, Sukarno N, Widyastuti U, Darusman LK, Kanaya S. Diversity of Endophytic Fungi from Red Ginger (*Zingiber officinale* Rosc.) Plant and Their Inhibitory Effect to *Fusarium oxysporum* Plant Pathogenic Fungi. *HAYATI Journal of Biosciences*. 2013; 20(3):127-137.
 50. Hanada RE, Pomella AWV, Costa S, Bezerra L, Loguercio L, Pereira J. O. Endophytic fungal diversity in *Theobroma cacao* (cacao) and *T. grandiflorum* (cupuaçu) trees and their potential for growth promotion and biocontrol of black-pod disease. *Fungal Biology*. 2010; 114(11-12):901-910.
 51. Louarn S, Nawrocki A, Thorup-Kristensen K, Lund OS, Jensen ON, Collinge DB, *et al.* Proteomic changes and endophytic micromycota during storage of organically and conventionally grown carrots. *Postharvest Biology and Technology*. 2013; 76:26-33.
 52. Siqueira VMde, Conti R, Araújo JMde, Souza-Motta CM. Endophytic fungi from the medicinal plant *Lippia sidoides* Cham. and their antimicrobial activity. *Springer*. 2011; 53:89-95.
 53. Redman RS, Kim YO, Woodward CJDA, Greer C, Espino L, Doty SJ, *et al.* Increased fitness of rice plants to abiotic stress via habitat adapted symbiosis: a strategy for mitigating impacts of climate change. *PLoS ONE*. 2011; 6.
 54. Qadri M, Johri S, Shah BA, Khajuria A, Sidiq T, Lattoo SK, *et al.* Identification and bioactive potential of endophytic fungi isolated from selected plants of the Western Himalayas. *SpringerPuls*. 2013; 2:8.
 55. Tomscheck AR, Strobel GA, Booth E, Geary B, Spakowicz D, Knighton B, *et al.* *Hypoxylon* sp., an endophyte of *Persea indica*, producing 1,8-cineole and other bioactive volatiles with fuel potential. *Microb Ecol*. 2010; 60(4):903-14.
 56. Ul-Hassan SR, Strobel GA, Booth E, Knighton B, Floerchinger C, Sears J. Modulation of volatile organic compound formation in the Mycodiesel-producing endophyte *Hypoxylon* sp. CI-4. *Microbiology*. 2012; 158(2):465-73.
 57. Numata A, Iritanj M, Yamada T, Minoura K, Matsumura E, Yamori T, *et al.* Novel antitumour metabolites produced by a fungal strain from a sea hare. *Tetrahedron letters*. 1997; 38(47):8215-8218.
 58. Sunayana N, Nalini MS, Kumara KKS, Prakash HS. Diversity studies on the endophytic fungi of *Vitex negundo* L. *Mycosphere*. 2014; 5(4):578-590.
 59. Markovskaja S, Kačergius A. Morphological and molecular characterization of *Periconia pseudobyssoides* sp. nov. and closely related *P. byssoides*. *Mycological Press*. 2014; 13(2):291-302.
 60. Prakash CP, Thirumalai E, Rajulu MBG, Thirunevukkarasu N, Suryanarayanan TS. Ecology and diversity of leaf litter fungi during early-stage decomposition in a seasonally dry tropical forest. *Fungal Ecology*. 2015; 17:103-113.
 61. Wong KM, Hyde KD. Diversity of fungi on six species of *Gramineae* and one species of *Cyperaceae* in Hong Kong. *Mycological Research*. 2001; 105(12):1485-1491.
 62. Marín P, Moretti A, Ritieni A, Jurado M, Vázquez C, González-Jaén MT. Phylogenetic analyses and toxigenic profiles of *Fusarium equiseti* and *Fusarium acuminatum* isolated from cereals from Southern Europe. *Food Microbiology*. 2012; 31(2):229-237.
 63. Tralamazza SM, Braghini R, Corrêa B. Trichothecene Genotypes of the *Fusarium graminearum* Species Complex Isolated from Brazilian Wheat Grains by Conventional and Quantitative PCR. *Frontiers in Microbiology (Online)*. 2016; 7:1.
 64. Rocha LO, Reis GM, Silva VN, Braghini R, Teixeira MMG, Corrêa B. Molecular characterization and fumonisin production by *Fusarium verticillioides* isolated from corn grains of different geographic origins in Brazil. *International Journal of Food Microbiology*. 2011; 145(1):9-21.
 65. Shweta S, Zuehlke S, Ramesha BT, Priti V, Kumar PM, Ravikanth G, *et al.* Endophytic fungal strains of *Fusarium solani*, from *Apodytes dimidiata* E. Mey. ex Arn (Icacinaceae) produce camptothecin, 10-hydroxycamptothecin and 9-methoxycamptothecin. *Phytochemistry*. 2010; 71(1):117-122.
 66. Amaral AL, Soglio FDK, Carli MLde, Neto JFB. Pathogenic fungi causing symptoms similar to *Phaeosphaeria* leaf spot of maize in Brazil. *Plant Disease*. 2005; 89:44-49.
 67. Costa IPdeMW. Fungos endofíticos isolados de vegetais do manguezal do rio Paripe, ilha de Tamaracá, Pernambuco, Brasil. Apresentado para a obtenção do grau de Mestre em Micologia Básica na Universidade Federal do Pernambuco, 2003.
 68. Kurose D, Furuya N, Tsuchiya K, Tsushima S, Evans C. Endophytic fungi associated with *Fallopia japonica* (Polygonaceae) in Japan and their interactions with *Puccinia polygoni-amphibii* var. *tovariae*, a candidate for classical biological control. *Fungal Biology*. 2012; 116(7):785-791.
 69. Sakalidis ML, Hardy GESJ, Burgess TI. Endophytes as potential pathogens of the baobab species *Adansonia gregorii*: a focus on the *Botryosphaeriaceae*. *Fungal Ecology*. 2011; 4(1):1-14.
 70. Jesus AEde, Steyn PS, Heerden FRvan, Vlegaar R,

- Wessels PL. Tremorgenic Mycotoxins from *Penicillium crustosum*: Isolation of Penitrems A-F and the Structure Elucidation and Absolute Configuration of Penitrem A. J. Chem. Soc. Perkin Trans. I 1983.
71. Gupta N, Das SJ. Phosphate Solubilising Fungi from Mangroves of Bhitarkanika, Orissa. Hayati Journal of Biosciences. 2008; 15(2):90-92.
 72. Acevedo E, Galindo-Castañeda T, Prada F, Navia M, Romero H M. Phosphate-solubilizing microorganisms associated with the rhizosphere of oil palm (*Elaeis guineensis* Jacq.) in Colombia. Applied Soil Ecology. 2014; 80: 26-33.
 73. Mouafi FE, Ibrahim GS, Elsoud MMA. Optimization of lovastatin production from *Aspergillus fumigatus*. Journal of Genetic Engineering and Biotechnology. 2016; 14(2):253-259.
 74. Bozza A, Tralamazza SM, Reynaud DT, Gabardo J, Valaski J, Marangoni PR, *et al.* Isolamento de fungos associados a grãos de café cv. Iapar 59 de origem de solo e árvore em diferentes tempos de colheita. Ciência e Tecnologia de Alimentos. 2009; 29:529-534.
 75. Almeida AB. Estratégias de controle e identificação de fungos produtores de ocratoxina A. Tese de doutorado, Setor de Patologia Básica, Microbiologia e Biologia Molecular, Acervo UFPR. 2015.
 76. Bussey III RO, Todd DA, Egan JM, El-Elimat T, Graf TN, Raja HA, *et al.* Comparison of the chemistry and diversity of endophytes isolated from wild-harvested and greenhouse-cultivated yerba mansa (*Anemopsis californica*). Phytochemistry Letters. 2015; 11:202-208.